



touchPANEL DISCUSSION

MET mutations: The next frontier in NSCLC testing

An expert panel discussion
recorded in July 2020

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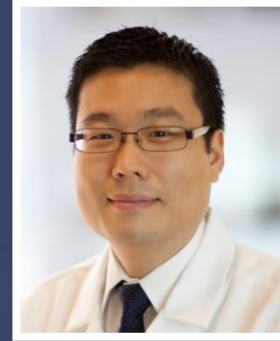
Expert panel



**Professor Karen Reckamp
(Chair)**
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Agenda

Mutation testing in NSCLC: who, what and when?

Presentation: Karen Reckamp

Panel discussion: Paul Paik and Keith Kerr; moderated by Karen Reckamp

How is molecular testing in NSCLC evolving?

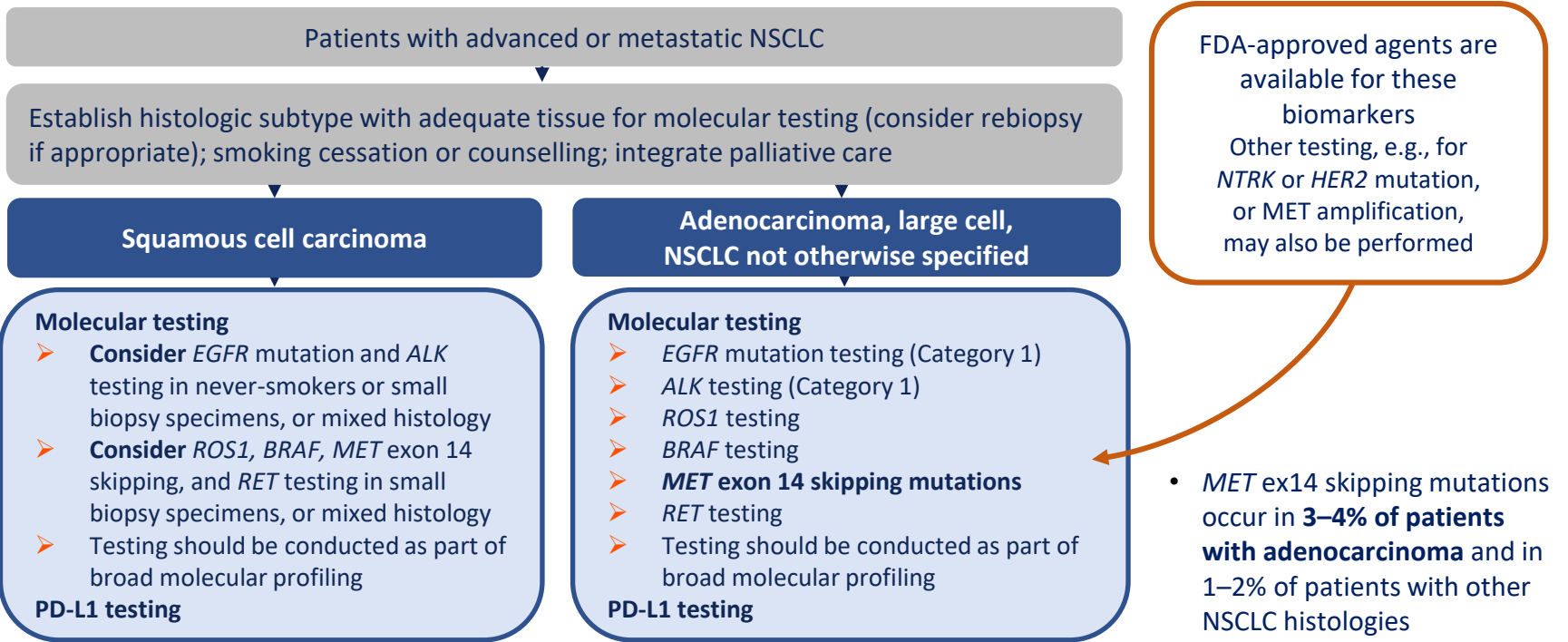
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Mutation testing in NSCLC: who, what and when?

NCCN guidelines for biomarker testing in NSCLC



ALK, anaplastic lymphoma kinase; *EGFR*, epidermal growth factor receptor; FDA, US Food and Drug Administration; *HER2*, human epidermal growth factor receptor 2; *MET* exon 14, mesenchymal–epithelial transition exon 14; NCCN, National Comprehensive Cancer Network; NSCLC, non-small cell lung cancer; *NTRK*, neurotrophic tropomyosin receptor kinase; PD-L1, programmed death-ligand 1; *RET*, rearranged during transfection.
NCCN Guidelines (Non-Small Cell Lung Cancer, Version 6.2020). Available at: www.nccn.org/professionals/physician_gls/pdf/nscl.pdf (accessed June 2020)

Timing of biomarker analysis in NSCLC



Before first-line therapy

- The NCCN panel emphasizes that molecular testing results for actionable biomarkers should be obtained before first-line therapy



At progression on targeted therapy

- Re-testing of a sample that is actively progressing while exposed to targeted therapy can shed light on appropriate next therapeutic steps

Other biomarker testing guidelines in NSCLC



Therapy-predictive biomarker testing in patients with metastatic NSCLC¹

- *EGFR* mutations
- *ALK* rearrangements
- *ROS1* rearrangements
- *BRAF* mutations
- PD-L1 expression

CAP/IASLC/AMP Guidelines²

- Include *MET*, *RET*, *HER2*, and *KRAS* in larger testing panels either initially or when negative for routine *EGFR*, *ALK*, *BRAF*, and *ROS1* testing

The National Lung Cancer Roundtable³

Guideline-concordant	Recommended	Optional as part of a larger panel	
<i>EGFR</i> , including <i>T790M</i>	MSI	<i>RET</i>	<i>KRAS</i>
<i>ALK</i>	PD-L1	<i>MET</i>	TMB
<i>BRAF</i>	<i>NTRK</i>	<i>HER2</i>	
<i>ROS1</i>			

ALK, anaplastic lymphoma kinase; AMP, Association for Molecular Pathology; CAP, College of American Pathologists; *EGFR*, epidermal growth factor receptor; FDA, US Food and Drug Administration; *HER2*, human epidermal growth factor receptor 2; IASLC, International Association for the Study of Lung Cancer; *MET* ex14, mesenchymal–epithelial transition exon 14; MSI, microsatellite instability; NGS, next-generation sequencing; NSCLC, non-small cell lung cancer; *NTRK*, neurotrophic tropomyosin receptor kinase; PD-L1, programmed death-ligand 1; *RET*, rearranged during transfection; TMB, tumor mutational burden.

1. ESMO Clinical Practice Guidelines, updated 18 September 2019. Available at: www.esmo.org/content/download/227453/3874538/1. (accessed June 2020);

2. Kalemkerian GP, et al. *J Clin Oncol*. 2018;36:911–9; 3. Kim ES, et al. *J Thorac Oncol*. 2019;14:338–42.

How is molecular testing in NSCLC evolving?

Tissue versus liquid biopsy testing in clinical practice



Tissue biopsy¹

- Historical standard of care
- Invasive; potential for bleeding and infection
- Difficulty obtaining adequate samples
- Not all patients suitable for biopsy
- Tumor DNA preserved in FFPE blocks
- Single-site tissue biopsies may miss genetic heterogeneity



Liquid biopsy (plasma ctDNA)^{1,2}

- Non-invasive; highly acceptable
- Potentially reduced cost and risk of complications
- An alternative when tissue biopsy specimens are insufficient or unfeasible
- Assesses DNA from all tumor sites; potentially bypasses intra-tumoral heterogeneity
- Can obtain serial samples at diagnosis and at acquired resistance
- Issues with sensitivity, specificity and standardization?


ctDNA, circulating tumor DNA; FFPE, formalin-fixed paraffin embedded.

1. Pennell NA, et al. *Am Soc Clin Oncol Educ Book*. 2019;39:531–42; 2. Rolfo C, et al. *J Thorac Oncol*. 2018;13:1248–68.

The evolving role of NGS testing in NSCLC

NGS is increasingly utilized in clinical laboratories¹

Benefits of NGS

- 
- ✓ Simultaneously tests for multiple alterations using a single tissue sample²
 - ✓ Should ultimately reduce costs and increase availability for patients, reducing the need for rebiopsy versus single-gene tests³
 - ✓ Single-gene test sequences are time-consuming and may require a relatively large tissue sample, which is not always available³

Challenges of implementing NGS

- Limited access, lack of awareness in medical care teams, limited coverage, and low reimbursement rates²
- Interpretation of NGS reports and use of results to guide treatment decisions³
- Careful consideration of limitations – e.g. not all assays that include *MET* will detect all known *MET* exon 14 skipping variants⁴

MET exon 14, mesenchymal–epithelial transition exon 14; NGS, next-generation sequencing; NSCLC; non-small cell lung cancer.

1. Ettinger DS, et al. *J Natl Compr Canc Netw*. 2018;16:807–821; 2. Pennell NA, et al. *J Clin Oncol*. 2018;36 (suppl; abstr 9031); 3. Pennell NA, et al. *Am Soc Clin Oncol Educ Book*. 2019;39:531–42; 4. Davies KD, et al. *J Thorac Oncol*. 2019;14:737–41.

DNA-based and RNA-based NGS testing in NSCLC

NGS panel assays can identify mutations, such as *MET* ex14 skipping events, to guide selection of targeted therapy^{1,2}



DNA sequencing²

- Detects genomic variants that alter or ablate a splicing site, or delete a whole exon



RNA sequencing²

- Detects the results of altered splicing (e.g. “fusion” of *MET* exon 13 to 15) regardless of the underlying genomic event
- RNA is more fragile than DNA, and high-quality RNA is harder to acquire from clinical cases



In a study of 286 samples:²

- RNA analysis detected *MET* ex14 skipping at 4.2% versus 1.3% with DNA analysis*
- Six of 10 positives by RNA analysis were not detected by DNA analysis
- RNA analysis was highly reliant on RNA quality

*DNA-based NGS assay using amplicon-mediated target enrichment; RNA-based NGS assay using anchored multiplex PCR for target enrichment.

MET, mesenchymal–epithelial transition; NGS, next-generation sequencing; PCR, polymerase chain reaction.

1. Qin D. *Cancer Biol Med.* 2019;16:4–10; 2. Davies KD, et al. *J Thorac Oncol.* 2019;14:737–41.



**Thank you for watching
this on-demand event**

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